Chemical composition and functional properties of two brown seaweeds from the Qeshm Island, Iran

Mohammadi E.^{1*}; Shabanpour B.¹; Kordjazi M.¹

Received: December 2016

Accepted: August 2017

Abstract

Seaweeds have been introduced as an important sources of food and pharmaceutical active agents since last century. In this study, the chemical composition and functional characteristics of (common names) *Nizamuddinia zanardinii* and *Iyengaria stellata*, were investigated. The total carbohydrate and ash contents were the two most abundant components in these seaweeds but their crude lipid and fiber contents were low. The protein content was higher in *I. stellata* followed over *N. zanardinii*. Atomic absorption spectrophotometry of the ashes showed that *I. stellata* contained higher amounts of both macrominerals (Ca, Mg) and trace elements (Fe, Zn) than *N. zanardinii*. Results of antioxidant properties showed that total phenolic (3.37 mg 100g⁻¹), total antioxidant (27.08 mg g⁻¹) and reducing power (22.76%) were higher in *N. zanardinii* than *I. stellata*, while DPPH free radical scavenging (7.16%) was higher in *I. stellata*. The results suggest that both the seaweeds could be used as a potential source of nutrient and antioxidant compounds in food and pharmaceutical industries.

Keywords: Brown seaweeds, Chemical composition, Antioxidant activity, Mineral composition

¹⁻Department of Fisheries, Faculty of Fisheries and Environmental Sciences, Gorgan University of Agricultural Sciences and Natural Resources, Gorgan, Iran *Corresponding author's Email: esmat.mohammadi69@gmail.com

Introduction

Seaweeds are marine large and diverse groups of photosynthetic organisms like plants which form the primary biomass in the coastal regions (Miyashita, 2009). About 9000 species of seaweeds have been identified and broadly classified into three principal classes based on their green pigmentation: (Chlorophytes), brown (Phaeophytes) and red (Rhodophytes) algae (Miyashita et al., 2013). Marine seaweeds are one of the living reproducible resources of the marine environment with potential food (Kumar et al., 2011) for human and animals, and medicine applications. Macroalgae are a rich source of polysaccharides, vitamins, antioxidant, dietary fiber, fatty acids, and minerals (Sinéad Lordan et al., 2011) and they are a potential source of secondary metabolites (Narayani et al., 2011). These bioactive compounds are a major resource for drug development (Kuda et al., 2005). In addition, marine macroalgae are used as sources of phycocolloids, thickening and gelling factors in food industries (Ruperez and Calixto, 2001). Algae also are a rich source of antioxidant compounds (Lim et al., 2002; Kuda et al., 2005; Chandini et al., 2008; Kokilam et al., 2013) and it has reported that brown been algae comparatively have higher antioxidant potential in comparison with green and red algae. (Kokilam et al., 2013). Polyphenols, carotenoids. tocopherols. terpenes. ascorbic acid and alkaloid are the natural compounds with antioxidant potential in seaweeds (Kokilam et al., 2013). These compounds react rapidly with radicals of oxygen such as hydroxyl, superoxide and peroxyl radicals are formed in human cells by endogenous and exogenous factors, and prevent from a wide range of human diseases (Chandini et al., 2008). In recent years, natural antioxidant compounds have attracted major interest as replace synthetic ones in prolonging the shelf life of food and cosmetics by delaying oxidation. In addition, biologically active compounds from marine algae have been proposed for nutraceuticals and functional foods (Balboa et al., 2013). Marine algae are distributed in the southern coast of Iran especially around seashores of Busheher and Hormuzgan provinces. Due to the presence of various species of seaweed in the Persian Gulf, reports on the antioxidant properties and chemical composition of seaweeds from Iran are very limited. Therefore, the current study was performed to measure the antioxidant properties and chemical composition of two various species of brown seaweeds. In this research, proximate composition, physicochemical properties, mineral content, total phenolic and antioxidant activity, DPPH free radical scavenging ability and reducing power of both Nizamuddinia zanardinii and Iyengaria stellata were evaluated. N. zanardinii and I. stellata (BØrgesen) are two various species of brown seaweeds with a similar class, Phaeophyceae, N. zanardinii belongs to the order Laminariales and family Alariaceae and I. stellata belongs to the order **Scytosiphonales** and family Scytosiphonaceae.

Materials and methods

Samples collection

Brown seaweeds *I. stellata* and *N. zanardinii* were collected from the coast of Qeshm Island in February 2014. The samples were washed thoroughly with

seawater to remove disturbing factors such as epiphytes and dirt particles, followed by shade-drying for two to five days. After that, the samples were transferred to the laboratory and the shade dried seaweeds were powdered to less than 1 mm particles in size. The milled seaweed samples were stored in sealed plastic bags at -20°C for further experiments.

Chemicals

Folin-Ciocalteau reagent, gallic acid. ammonium molybdate, ascorbic acid. buffer. Sodium phosphate carbonate. Iron(III) chloride-6-hydrate, Trichloroacetic acid. 1,1-diphenyl-2picrylhydrazyl (DPPH), and potassium ferricyanide, were purchased from sigma-Aldrich chemical co. Inc. (St. Louis, Mo, USA).

Estimation of proximate composition of the seaweed samples

Ash content

The ash content of seaweeds was quantified gravimetrically according to the method of Seedevi *et al.* (2015). The ash content was expressed as percent of dry weight.

Crude fiber content

Fiber quantified using the Fibertec System (IKA, Germany) (Robledo and Freile Pelegrín, 1997). The fiber content of the samples reported as percent of dry weight.

Extractable fat

Determining the fat percentage was done by Soxhlet method and by using petroleum ether solvent (Peinado *et al.*, 2014).

Protein

Total protein was determined by the micro-Kjeldahl method. The protein was specified by using a nitrogen conversion coefficient of 6.25 (Akgül *et al.*, 2015). Data reported as a percentage of dry weight.

Moisture content

The moisture content of the seaweeds was determined by drying following method of Seedevi *et al.* (2015).

Estimation of total carbohydrate content

Estimation of total carbohydrate was conducted by Phenol-sulphuric acid method (Syad *et al.*, 2013). The total carbohydrate content of seaweeds was estimated based on Standard glucose curve and the results were expressed in percentage.

Evaluation of mineral contents

Mineral contents were quantified using the atomic absorption spectrophotometry (Perkin-Elmer Analyst 800 with flame furnace) (Syad *et al.*, 2013). The minerals analyzed were Calcium, Iron, Magnesium, Manganese, Copper, Cadmium, Lead, and Zinc and the results were expressed as mg/100g dry weight.

Preparation of samples extract for antioxidant activities

Extraction was conducted in three steps. The extraction of seaweed sample was performed with distilled water. Briefly, 50 grams of the algal powder was mixed with 1000 ml of distilled water, were shaken in an incubator shaker (IKA® KS 4000 ic control, Germany) at room temperature for 24 h at 200 rpm and the mixture was centrifuged at 3500 rpm (5810R, Eppendorf, Germany) for 10 min at 4°C and filtered with Whatman no 4 filter paper. Both concentrate and supernatant were freeze-dried and the concentrate was weighed. At the secondary and third steps, distilled water was added to the concentrate at the ratio of 1:20 (W/V) and the above process was repeated. The dried triplicate extracts were pooled and stored at -80°C until analyzed. Each dried extraction was then dissolved in distilled water at a concentration of 5mg/ml as a stock solution. The stock solution was applied both for the determination of TPC and antioxidant activities.

Total phenolic content (TPC)

TPC in the aqueous extracts was assessed with Folin-Ciocalteu reagent based on the method of Taga et al. (1984). Various concentrations of 0.001 mg ml⁻¹ to 1.0 mg ml⁻¹ of standard stock solution were prepared. Briefly, 200 ml of 2% Na₂CO₃ was mixed with 100 µl of test solutions. After 2 min, test solutions were mixed with 100 µl of 50% Folin-Ciocalteau reagent and were placed at temperature 25°C for 30 min. Spectrophotometer (Biochrom, England) was used to read the absorbance value at 750 nm. Gallic acid was used as a standard. Total phenolic content was assessed by comparison with the standard calibration curve and the results were expressed as milligram Gallic acid equivalent per 100 gram of extract.

Total antioxidant activity

Total antioxidant activities of aqueous extracts were assessed based on the method of Sathya *et al.* (2013). Briefly, 0.3 mL of test sample solution at different concentrations (10-100 mg ml⁻¹) was added to 3 mL of reagent (0.6 M sulfuric acid, 28 mM sodium phosphate and 4 mM ammonium molybdate). The mixture was placed in a Ben Murray with temperature 95°C for 90 min. Then the absorbance was read at 695 nm. Ascorbic acid was used as standard and total antioxidant capacity reported as milligram ascorbic acid equivalent per gram of extract.

DPPH scavenging activity

The scavenging effects of samples for DPPH radical were monitored based on the method of Rodríguez-Meizosoet et al. (2006). Briefly, 23.5 mg of DPPH powder was dissolved in 200 ml of methanol. This solution was diluted with methanol at a ratio of 1:20; then 0.1 ml of different concentrations of algal extracts (20-200 µg ml⁻¹) added to 3.9 ml of a methanolic solution of DPPH. The test solutions were vortexed for 1 min and kept in the dark at 25°C for 30 min. The absorbance of all the sample solutions was measured at 517 nm by using the methanol as blank. Synthetic antioxidants Gallic acids and Ascorbic acid were used as positive controls. The DPPH free radical scavenging ability was estimated using the following formula: Scavenging effect (%)= $[(A_0-A_1)/A_0] \times 100$ A₀=Absorbance of control; A₁=Absorbance of sample

 A_0 =DPPH solution without sample; A_1 =DPPH solution contain test sample

Reducing power

The reducing power of aqueous extracts was assessed based on the method of Kumar *et al.* (2011). Briefly, 1.25 ml of phosphate buffer (0.2 M, pH 6.6) and 1.25 ml of potassium ferricyanide (1%) were

mixed with 0.5 ml of extract samples solution at concentrations (10-320 ug ml⁻¹) and the mixture was incubated at 50°C for 20 min. Afterward 1.25 ml of trichloroacetic acid (10%), 1.25 of ml distilled water and 0.25ml of Fecl3.6H2O (0.1%) were added to the mixture. The mixture was kept at 25 °C for 10 min and the absorbance was measured at 700 nm. Synthetic antioxidant Ascorbic acid was used as positive control. Reducing power increases by increasing the absorbance value of the reaction mixture.

Statistical analysis

All analyses conducted in triplicates, the data reported as means±standard deviations. Statistical analysis was performed by software SPSS (Version 20).

Independent samples t-test was implemented to assess for any significant differences between the means. pvalue<0.05. *p*<0.01 regarded as significant. Microsoft Office Excel 2010 used to diagramming.

Results

The proximate composition of Ν. zanardinii and I. stellata were investigated and the results were tabulated in Table 1. The amounts of ash and protein in I. stellata. moisture and fiber were significantly higher in N. zanardinii (p < 0.05). But the fat and carbohydrate contents of algae N. zanardinii and I. stellata were not observed a statistically significant difference (p>0.05).

Table 1: Proximate com	position of the l	Nizamuddinia zana	<i>rdinii</i> and <i>l</i>	yengaria stellata
------------------------	-------------------	-------------------	----------------------------	-------------------

Composition	N. zanardinii	I. stellata
Ash content	15.84 ± 1.26^{b} % DW	33.68 ± 3.25^{a} % DW
Crude fiber content	$5.18 \pm 0.07^{a} \% DW$	$4.68 \pm 0.08^{b} \% \text{ DW}$
Crude protein content	3.18 ± 0.05^{b} % DW	$8.16 \pm 0.14^{a} \% DW$
Crude lipid content	$0.41 \pm 0.12^{a} \% DW$	0.27 ± 0.09^{a} % DW
Moisture content	$8.99 \pm 0.17^{a} \% \text{ DW}$	3.77 ± 0.67^{b} % DW
Total carbohydrate content	29.10 ± 5.60^{a} % DW	$34.80 \pm 5.41^{a} \% DW$

Results expressed as Mean \pm SD (n= 3). Different superscript letters in the same row indicate significant differences (p<0.05). DW: dry weight.

Mineral composition

The mineral composition of two different species of brown seaweeds was shown in Table 2. There were significant (p<0.05) differences in the concentrations of all the mineral and trace elements except Magnesium and Copper between two species of seaweeds. Among all the 8 minerals analyzed Calcium were found to be highest (2580-9586 mg $100g^{-1}$ DW) in *N. zanardinii* and *I. stellata*, respectively. The Lead was a toxic element with the lowest concentration in the analyzed seaweeds, the range of <0.5mg $100g^{-1}$ - <0.5mg $100g^{-1}$ for the *N. zanardinii* and *I. stellata*, respectively.

Mineral	Seaweeds		
	N. zanardinii	I. stellata	
Ca	2580 ± 278.74^{b}	$9586.66 \pm 476.069^{\rm a}$	
Mg	1316.66 ± 32.14^{a}	1316.66 ± 25.16^{a}	
Fe	$90 \pm 10^{\mathrm{b}}$	326.66 ± 15.27^{a}	
Mn	$2.06\pm0.49^{\rm b}$	14.43 ± 0.30^{a}	
Zn	$1\pm0.1^{ m b}$	$1.53\pm0.05^{\mathrm{a}}$	
Cu	$0.73\pm0.57^{\rm a}$	$0.53\pm0.05^{\mathrm{a}}$	
Pb	<0.5	<0.5	
Cd	<0.2	0.2 ± 0	

 Table 2: Mineral composition (mg 100g⁻¹ dry weight) determined by atomic absorption spectrophotometry in Nizamuddinia zanardinii and Iyengaria stellate.

Results expressed as Mean \pm SD (n= 3). Different superscript letters in the identic row indicate significant differences (p<0.05).

Phenolic content

The total phenolic content (TPC) of *N. zanardinii* and *I. stellata* shown in Figure1. The phenolic content in extracts was significantly different between species

(p<0.05). The aqueous extract of *N*. *zanardinii* showed higher total phenolic content (TPC) of 3.37±0.13 mg GAE 100g⁻¹ compared to water extract of *I*. *stellata* 1.22±0.09 mg GAE 100g.



Figure 1: Total phenolic content of seaweeds extract. Different superscripts letters indicate significant differences (*p*<0.05). Data expressed as Mean±SD (n= 3).

Total antioxidant activity

Total antioxidant activity (TAE) is a method for determining the antioxidant potential of marine algae. In the present study, *N. zanardinii* and *I. stellata* showed the total antioxidant activity in the range of 0-27.08 mg g⁻¹ and 0-5.03 mg g⁻¹, respectively at different concentrations

(10-100 mg ml⁻¹). The maximum of 27.08 mg g⁻¹ and 5.03 mg g⁻¹ inhibition were observed at the concentration of 100 mg ml⁻¹, respectively, it can be concluded, By increasing the concentration of the extract, antioxidant activity increases. At concentrations of 10, none of the seaweeds showed antioxidant activity, but in other

concentrations the antioxidant activity of *N. zanardinii* was significantly higher than





Figure 2: Total antioxidant activity of seaweeds extract. Different superscript letters in the identic concentration indicate significant differences (p<0.05). Data expressed as Mean±SD (n=3).

The DPPH free radical scavenging

Antioxidant activities of the aqueous extracts of two species of brown seaweed were measured by using a radical scavenging method. In the present study inhibitory effect of extracts of Ν. zanardinii and I. stellata on DPPH radical are determined and depends on the $20-200 \ \mu g \ ml^{-1}$ concentration. The concentration showed 1.61-2.73% and 6.04-7.16% of inhibition respectively. The highest value of 2.73% and 7.16% observed at the concentration of 200 µg ml⁻¹ in N. zanardinii and I. stellata, respectively. The results showed that by

increasing the concentration of seaweed extracts the percentage of DPPH free radical scavenging increased. At the highest concentration, the standards of Gallic acid and ascorbic acid showed the antioxidant activity of 95.52-87.36% respectively. Inhibitory effect of positive controls in all concentrations compared to algae extracts were significantly higher (p < 0.01). DPPH free radical scavenging ability of an aqueous extract of brown algae I. stellata in all concentrations was significantly higher than N. zanardinii (*p*<0.05).



Figure 3: DPPH antioxidant activity of seaweeds extract. Different superscript letters in the identic concentration indicate significant differences (p<0.05). Different superscript emblems (++,--) in the same concentration indicate important significant differences with ascorbic acid and Gallic acid, respectively (p<0.01). Data expressed as Mean±SD (n= 3).

Reducing power

Fig. 4 shows the reducing power results of seaweeds *N. zanardinii* and *I. stellata* exhibited concentration dependent. The reducing power of *N. zanardinii* and *I. stellata* was 0.56-22.76% and 0.2-6.4% at the concentration of 10-320 μ g ml⁻¹ respectively, which showed a constant increase in its reducing power with increased concentration. Between the

extracts in concentrations of 20 and 40 were not observed statistically significant (p>0.05).in difference But other concentrations reducing power of macroalgae N. zanardinii was significantly higher (p < 0.05). Reducing power of ascorbic acid at all concentrations was significantly higher than the algae extracts (*p*<0.01).



Figure 4: Reducing power of Nizamuddinia zanardinii and Iyengaria stellate. Different superscript letters in the identic concentration indicate significant differences (p<0.05). Different superscript emblem (++) in the same concentration indicate important significant differences with ascorbic acid (p<0.01). Results are expressed as Mean±SD (n= 3).

Discussion

The Proximate composition of the N. zanardinii and I. stellata was shown in table 1. In general, the amount of ash is high in the seaweeds (ranges of 8 to 40% dry weight) (Rupérez, 2002) that is the indicative of high levels of various minerals in the seaweeds (Kumar et al., 2011). The ash content determined for I. stellata was higher than reported by Rohani-Ghadikolaei et al. (2012) and Syad et al. (2013). The total fiber content of seaweeds indicated a significant correlation (p<0.05). N. zanardinii and I. stellata have a higher fiber than nodosum, Caulerpa Ascophyllum racemosa, Codium isthmocladum and Eucheuma isiforme, but the fiber was lower in these seaweeds when compared to Sargassum filipendula and Padina gymnospora (Robledo and Freile Pelegrín, 1997). In general, total fiber content was low in both seaweeds. The protein values are significantly different between the two seaweeds (p < 0.05). The amount of protein in brown seaweeds is within the ranges of 3 to 15% of dry weight of seaweed (Burtin, 2003; Chakraborty and Santra, 2008; Manivannan et al., 2008, 2009; Rohani-Ghadikolaei et al., 2011). Variations in the protein content of seaweeds can occur due to species, season and their environmental conditions (Dawczynski et al., 2007). Lipids are one of the components of seaweeds. The lipid level of seaweeds is generally low (1-5% of dry weight) (Burtin, 2003; Polat and Ozogul, 2008). The levels of lipids detected in the present study were more than previously reported for other brown seaweed species, namely: Sargassum wighti (0.21±0.001%) and Hormophysa

triquetra (0.11±0.001%), (Kokilam et al., 2013). Carbohydrates are needed for important energy processes such as (Kokilam al.. respiration et 2013). Fucoidan, laminaran, cellulose, alginate mannitol are the and common in carbohydrates brown seaweeds (Dawczynski et al., 2007). The carbohydrate values are not significantly different between the two seaweeds (p>0.05). These results are higher than those reported by MacArtain et al (2007) for Ascophyllum nodosum (13.1%),Laminaria digitata (9.9%), Himanthalia elongata (15%), Undaria pinnatifida (4.6%), Mohammadi *et al* (2012) for Colpomenia sinuosa (11.3%), and Syad et al (2013) for S. wightii (0.0095%). Environmental factors such as temperature, salinity and sunlight intensity can change the amount of carbohydrates in different species (Sakthivel and Devi, 2015). Moisture is an important factor in determination of the shelf-life and quality of food materials (Syed et al., 2013). was a significant There correlation between moisture in the seaweeds (p < 0.05).

Atomic absorption spectrophotometry determination of the ashes showed that these seaweeds contained high amounts of the macrominerals and trace elements are required in human diet. I. stellata contains more minerals due to having more ash content. Differences in the amount and composition of polysaccharides in cell walls of seaweeds could be explained due to differences in the biosorption of minerals and trace elements (Davis et al., 2003; Bocanegra et al., 2009). The Iron, Magnesium and Copper content determined for seaweeds in this study were

higher than brown seaweeds reported by al.Krishnaiah et (2008): Rohani-Ghadikolaei, et al. (2012) and Astorga-España et al. (2015). The Cadmium, Lead, Copper and Zinc consumption would be 3.77, 0.91, 10.79 and 25.35 μ g day⁻¹, respectively which FAO/WHO expert committee has allowed these values of elements for daily consumption. Copper and Zinc content in N. zanardinii and I. stellata were lower than reported limits by FAO/WHO (Subba Rao et al., 2007).

Phenolic compounds are commonly found in plants but earlier reports revealed that marine seaweed extracts possess antioxidant properties (Lim et al., 2002). Antioxidants phenolic are the most effective compounds in brown seaweeds 2005). et al., Polyphenols, (Kuda phlorotannins, and fucoxanthin are the major active compounds in different seaweed extracts that contain several biological activities including antioxidant properties (Yan et al., 1996). Total phenol content in N. zanardinii studied in this work was lower than N. zanardinii reported from Chabahar coast (Fariman et al., 2015). Further, phenol content in I. stellata tested in this research was lower than I. stellata reported from Bandar Moalam (Moein et al., 2015).

Natural antioxidants are not limited to terrestrial sources and reports show that seaweeds are rich sources of natural antioxidant compounds (Cox *et al.*, 2010). Total antioxidant activity in *N. zanardinii* is higher than those reported by Kokilam *et al.* (2013) for *Hormophysa triquetra* (24.0 \pm 3.05 mg g⁻¹) and *Sargassum wightii* (20.0 \pm 2 mg g⁻¹).

DPPH is a useful reagent for evaluation of free radical-scavenging activities of compounds and it accepts an electron or hydrogen radical to become a stable diamagnetic molecule. When DPPH is mixed with a substrate acting as a hydrogen atom donor, a stable non-radical form of DPPH is formed, with the simultaneous change in the color of the solution from violet to pale yellow (Sathya et al., 2013). DPPH radical scavenging activity of the positive controls, Gallic acid, and ascorbic acid, was higher than water extract of the N. zanardinii and I. stellata in all of five tested concentrations. DPPH radical scavenging activity in I. stellata was higher compared to Nizamuddinia zanardinii and Cystoseira indica (Fariman et al., 2015). But DPPH radical scavenging in N. zanardinii tested this research was lower than in Nizamuddinia zanardinii reported from Chabahar coast (Fariman et al., 2015).

In the reaction system, antioxidant substances in the samples cause reduction of the Fe³⁺/ferricyanide complex to the Fe^{2+} form by breaking the free radical chain by donating a hydrogen atom. The reaction of reductions with special precursors of peroxide leads to prevention of the formation of peroxide (Seedevia et al., 2015). Thus, the antioxidant activity of an aqueous extract related to the reductive activity. The increased absorbance value of samples indicates increased reducing power. Higher absorbance value means stronger reducing the power of sample. Reducing power in N. zanardinii and I. stellata were higher than Colpomenia sinousa, Cystoseria myrica and Iyengaria stellata (Moein et al., 2015).

Seaweeds have been introduced as important sources of food and pharmaceutical active agents since last century. The two seaweeds analyzed for their chemical composition were found to be interesting potential sources of carbohydrate and minerals owing to their high levels. Protein content was high in I. stellata. Results of antioxidant properties showed that total phenolic content, total antioxidant activity and reducing power were high in N. zanardinii than the I. while DPPH free stellata. radical scavenging was higher in I. stellata. So, these seaweeds can be considered as potential sources of natural antioxidants. Persian Gulf seaweeds studied can be useful as a substitute for synthetic antioxidants, medical applications or as additives. The findings of this work can be helpful to future research to identify bioactive antioxidant compounds.

Acknowledgments

The authors thank the chemistry laboratory of Gorgan University of Agricultural Sciences and Natural Resources and also the Fishery Research Center of Qeshm for their technical support.

References

- Akgül, R., Kızılkaya, B., Akgül, F. and Erduğan, H., 2015. Amino acid composition and crude protein values of some Cyanobacteria from Çanakkale (Turkey). *Pakistan Journal of Pharmaceutical Sciences*, 28(5), 1757-1761.
- Astorga-España, M.S., Rodríguez
 Galdón, B., Rodríguez
 Rodríguez
 Rodríguez, Rodríguez,
 E.M. and Díaz
 Romero, C., 2015.
 Mineral and trace element concentrations in seaweeds from the sub-Antarctic ecoregion of Magallanes

(Chile). Journal of Food Composition and Analysis, 39, 69-76.

- Balboa, E.M., Conde, E., Moure, A., Falqué, E. and Domínguez, H., 2013. In vitro antioxidant properties of crude extracts and compounds from brown algae. *Food chemistry*, 138(2), 1764-1785.
- Bocanegra, A., Bastida, S., Benedí, J., Ródenas, S. and Sánchez-Muniz, F.J., 2009. Char-acteristics and nutritional and cardiovascular-health properties of seaweeds. *Journal of Medicinal Food*, 12, 236-258.
- Burtin, P., 2003. Nutritional value of seaweeds. *Electronic Journal of Environmental, Agricultural and Food Chemistry*, 2(4), 498-503.
- Chakraborty, S. and Santra, S.C., 2008. Biochemical composition of eight benthic algae collected from Sunderban. *Indian Journal of Marine Sciences*, 37(3), 329.
- Chandini, S., Ganesan, P. and Bhaskar, N., 2008. In vitro antioxidant activities of three selected brown seaweeds of India. *Food Chemistry*, 107, 707-713.
- Cox, S., Abu-Ghannam, N. and Gupta, S., 2010. An assessment of the antioxidant and antimicrobial activity of six species of edible Irish seaweeds. *International Food Research Journal*, 17, 205-220.
- Davis, T.A., Volesky, B. and Mucci, A., 2003. A review of the biochemistry of heavy metal biosorption by brown algae. *Water Research*, 37, 4311-4330.
- Dawezynski, C., Schubert, R. and Jahreis, G., 2007. Amino acids, fatty acids and dietary fiber in edible seaweed products. *Food Chemistry*, 103, 891-899.

- Fariman. **G.A.**, Shastan, S.J. and M.M., 2016. Seasonal Zahedi. variation of total lipid, fatty acids, fucoxanthin content, and antioxidant properties of two tropical brown algae (Nizamuddinia zanardinii and Cystoseira indica) from Iran. Journal of Applied Phycology, 28(2), 1323-1331.
- Kokilam, G., Vasuki, S. and Sajitha, N., 2013. Biochemical composition, alginic acid yield and antioxidant activity of brown seaweeds from Mandapam region, Gulf of Mannar. *Journal of Applied Pharmaceutical Science*, 3(11), 99.
- Krishnaiah, D., Sarbatly, R., Prasad, D.M.R. and Bono, A., 2008. Mineral content of some seaweeds from Sabah's South China Sea. Asian Journal of Scientific Research, 1, 166-170.
- Kuda, T., Tsunekawa, M., Goto, H. and Araki, Y., 2005. Antioxidant properties of four edible algae harvested in the Noto Peninsula, Japan. *Journal of Food Composition and Analysis*, 18, 625-633.
- Kumar, M., Gupta, V., Kumari, P., Reddy, C.R.K. and Jha, B., 2011. Assessment of nutrient composition and antioxidant potential of Caulerpaceae seaweeds. *Journal of Food Composition and Analysis*, 24, 270-278.
- Lim, S.N., Cheung, P.C.K., Ooi, V.E.C. and Ang, P.O., 2002. Evaluation of antioxidative activity of extracts from a brown seaweed, Surgassum siliquastrum. *Journal of Agricultural and Food Chemistry*, 50, 3862-3866.
- MacArtain, P., Gill, C.I., Brooks, M., Campbell, R. and Rowland, I.R., 2007. Nutritional value of edible seaweeds. *Nutrition Reviews*, 65(12), 535-543.

- Manivannan, K., Thirumaran, G., Devi, G.K., Hemalatha. A. and Anantharaman, P., 2008. Biochemical of composition seaweeds from Mandapam coastal regions along Southeast Coast of India. American-Eurasian Journal of Botany, 1(2), 32-37.
- Manivannan, K., Thirumaran, G., Devi, Anantharaman, G.K.. **P**. and Balasubramanian, Т., 2009. Proximate composition of different group of seaweeds from Vedalai Coastal waters (Gulf of Mannar): Southeast Coast of India. Middle-East Journal of Scientific Research, 4(2), 72-77.
- Miyashita, K., 2009. The carotenoid fucoxanthin from brown seaweed affects obesity. *Lipid Technology*, 21(8-9), 186-190.
- Miyashita, K., Mikami, N. and Hosokawa, M., 2013. Chemical and nutritional characteristics of brown seaweed lipids: A review. *Journal of Functional Foods*, 5, 1507-1517.
- Moein, S., Moein, M., Ebrahimi, N., Farmani, F., Sohrabipour, J. and Rabiei, R., 2015. Extraction and determination of protein content and antioxidant properties of ten algae from Persian Gulf. *International Journal of Aquatic Science*, 6(2), 29-38.
- Mohammadi, M., Tajik, H. and Hajeb,
 P., 2013. Nutritional composition of seaweeds from the Northern Persian Gulf. *Iranian Journal of Fisheries Sciences*, 12(1), 232-240.
- Narayani, C.G.S., Arulpriya, M., Ruban, P., Anantharaj, K. and Srinivasan, R., 2011. In vitro antimicrobial activities of seaweed

extracts against human pathogens. Journal of Pharmacy Research, 4(7), 2076-2077.

- Parthiban, C., Saranya, C., Girija, K., Hemalatha, A., Suresh, M. and Anantharaman, P., 2013. Biochemical composition of some selected seaweeds from Tuticorin coast. Advances in Applied Science Research, 4(3), 362-366.
- Peinado, I., Girón, J., Koutsidis, G. and Ames, J.M., 2014. Chemical composition, antioxidant activity and sensory evaluation of five different species of brown edible seaweeds. *Food Research International*, 66, 36-44.
- Polat, S. and Ozogul, Y., 2008. Biochemical composition of some red and brown macro-algae from the Northeastern Mediterranean Sea. *International Journal of Food Sciences and Nutrition*, 59, 566-572.
- Robledo, D. and Pelegrín, Y.F., 1997. Chemical and mineral composition of six potentially edible seaweed species of Yucatan. *Botanica Marina*, 40, 301-306.
- Rodríguez-Meizoso, I., Marin, F.R., Herrero, M. and Señorans, F.J., 2006. Subcritical water extraction of nutraceuticals with antioxidant activity from oregano. Chemical and functional characterization. *Journal of Pharmaceutical and Biomedical Analysis*, 41, 1560-1565.

Rohani-Ghadikolaei, K.,

Abdulalianand, E. and Ng, W.K., 2012. Evaluation of the proximate, fatty acid and mineral composition of representative green, brown and red seaweeds from the Persian Gulf of Iran as potential food and feed resources. Journal of Food Science and Technology, 49, 774-780.

- Rupérez, P. and Calixto, F.S., 2001. Dietary fiber and physico-chemical properties of edible Spanish seaweeds. *European Food Research and Technology*, 212, 349-354.
- Rupérez, P., 2002. Mineral content of edible marine seaweeds. *Food Chemistry*, 79, 23-26.
- Sakthivel, R. and Devi, K.P., 2015. Evaluation of physicochemical properties, proximate and nutritional composition of *Gracilaria edulis* collected from Palk Bay. *Food Chemistry*, 174, 68-74.
- Sathya, R., Kanaga, N., Sankar, P. and Jeeva, S., 2013. Antioxidant properties of phlorotannins from brown seaweed Cystoseira trinodis (Forsskål) C. Agardh. Arabian Journal of Chemistry, 10, 2608-2614.
- Seedevia, P., Moovendhana, M., Sudharsana, S., Vasanthkumarb, Sh., Srinivasanc, A., Vairamania, Sh. and Shanmugamaa, A., 2015. Structural characterization and bioactivities of sulfated polysaccharide from Monostroma oxyspermum. International Journal of Biological Macromolecules, 72, 1459-1465.
- Subba Rao, P.V., Mantri, V.A. and Ganesan, K., 2007. Mineral composition of edible seaweed *Porphyra vietnamensis. Food Chemistry*, 102, 215-218.
- Sinéad Lordan, R., Ross, P. and Stanton, C., 2011. Marine Bioactives as Functional Food Ingredients: Potential to Reduce the Incidence of Chronic Diseases. *Marine Drugs*, 9, 1056-1100.

- Syad, A.N., Shunmugiah, K.P. and Kasi, P.D., 2013. Seaweeds as nutritional supplements: Analysis of nutritional profile, physicochemical properties and proximate composition of G. acerosa and S. wightii. *Biomedicine and Preventive Nutrition*, 3, 139-144.
- Taga, M.S., Miller, E.E. and Pratt, D.E., 1984. Chia seeds as a source of natural lipid antioxidants. *Journal of the American Oil Chemists' Society*, 61(5), 928-931.
- Yan, X.J., Li, X.C., Zhou, C.X. and Fan, X., 1996. Prevention of fish oil rancidity by phlorotannins from Sargassum kjellmanianum. *Journal of Applied Phycology*, 8, 201-203.